Synergy Effect of Aspergillus Terreus and Aspergillus Fumigatus in Bioremediation of Chromium from Tannery Effluent

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Abstract- Large volume of wastewater is generated by tannery industry, which contains heavy metals such as Chromium, Cadmium, Lead, Copper, Zinc, Mercury and Arsenic. Chromium (Cr) is one of the common and toxic heavy metal pollutants released by industries in their effluent. In this study, tannery effluent was collected from Challawa industrial estate, Kano, Nigeria. Physico-chemical parameters of the effluent were determined; two fungal isolates Aspergillus terreus and Aspergillus fumigatus were obtained from the wastewater. The industrial effluent was supplemented with minimal salts medium: 3 mg/L NH₄Cl, 3 mg/L K₂HPO₄, 5 mg/L KH₂PO₄, 1 mg/L NaCl, 1mg/L MgSO₄ and tyndallized. The Cr content in effluent were measured before inoculation of the fungal isolates and after using Atomic Absorption Spectrophotometer (Model GBC 932). The co-isolates were found to absorb 97.32% of Cr in the effluent. Scan Electron Microscopy (SEM) was also carried out. Bioremediation is an eco-friendly and economically feasible process.

Keywords- Synergy, Aspergillus terreus, Aspergillus fumigatus Bioremediation, Chromium

I. INTRODUCTION

Tannery industry is an important activity in many developing countries, it has been estimated that about 18 feet square of leather are made annually around the world with huge amount of many of money involved. This industry uses large quantities of water and in turn produces large amount of effluents [1]. In leather tanning, a number of processes are performed using large number chemicals such as acids, surfactants, dyes, natural or synthetic tanning agents, sulfonated oils and salts to convert animals skins into leather products [2]. The effluent contain low biodegradable chemicals, tannery wastewaters cause serious environmental problems. In addition, the effluent contains high concentrations of chemical oxygen demand (COD), dissolved oxygen (DO), biological oxygen demand (BOD₅) and total suspended solids (TDS), which leads to persistent of the pollutants in the effluent [3]. Chromium salts have a high rate of penetration into the interfibrillar spaces of the skins. In spite of that, only a fraction of the salts used in the tanning process yields the desired reaction with the skins [4]. This happens because of the limited access to reactive sites for the tanning molecules, and/or the presence of low affinity species in the tanning materials. The rest of the salts remain in the tanning effluents and are subsequently sent to a treatment plant where the Chromium salts end up in the waste Chromium is the second most abundant inorganic groundwater contaminant at hazardous waste sites. It was observed that Cr (III) readily oxidizes to Cr(VI) when typical oxidation conditions are present [5]. The toxicity of Cr depends primarily on its chemical form. Trivalent Chromium compounds are much less toxic than those of hexavalent Cr, since hexavalent Chromium is known to be very mobile and hazardous to human health through inhalation, skin contact and ingestion. It is highly toxic, carcinogenic and mutagenic to living organisms, even when at very low concentrations in water [6]. The conventional methods of treatment such as coagulation, flocculation, advanced oxidation, ozonation and activated adsorption are not economically feasible and efficient for the heavy metal removal. The biological method of treatment of wastewater was found more effective and less costly, in which microbes are used for pollution control technology, the biological system are used to degrade or transform of various toxic chemicals to less harmful forms [7]. It mainly shows interest towards the pollution control using bacteria, fungi in combination with physicochemical methods [8, 9]. The fungal biomass has more advantage in absorbing heavy metals due their nature of mycelial growth and this makes them to reduce the

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levels of metal ion in wastewater. The biological treatment is cost effective and cleanup process of the pollutants with no harmful effect of secondary metabolites [7].

II. MATERIALS AND METHODS

In the preparation of reagents chemicals of analytical Analar grade were used with deionised water. All glass ware were cleaned and rinsed with detergents and immersed in 25% nitric acid and finally rinsed with de ionized water as conducted by [10]. Samples were collected from Global and Harmatan tanneries industries in the Challawa industrial estate, Kano State, Nigeria. The Samples were collected in triplicate in sterile bottles for physicochemical parameters, heavy metal and microbiological analyses. The pH of the effluent was measured at the point of collection; samples were stored at 4 C and transported to the laboratory for further investigations [11].

II.I LOCATION OF THE STUDY AREA

Challawa industrial estate is located in Kumbotso Local Government Area of Kano State. It is located in the northern Nigeria covering an area extending between latitude 12° 40' and 10° 30' and longitude 7° 40' and 90° 40'. The industries in the Challawa industrial estate range from tanneries and textiles to food and packaging / processing [12, 13].

II.II ANALYSIS OF PHYSICOCHEMICAL PARAMETERS

The physicochemical parameters were measured before and after treatment with the fungal isolates. The parameters include the following, pH, DO, COD, BOD₅, TSS, TDS, Turbidity and Electrical Conductivity [11].

II.III ISOLATION AND IDENTIFICATION OF FUNGAL ISOLATES

Serial dilution of the effluent was carried out and 1 mL was put on the Sabouraud Dextrose Agar (SDA), incubated at 37°C for 4 - 5 days. Working solution of 100 - 400 mg/L of K₂CrO₄ was prepared. The solutions were enriched with Sabouraud Dextrose Broth (SDB), sterilized and inoculated, kept at 37°C in the orbital shaker at 100 rpm for 5 days [14]. The two isolates were able to grown at 400 mg/L concentration of K₂CrO₄ were picked as the potential isolates for bioremediation. The morphology of isolate was studied by lacto phenol cotton blue staining [15, 16, 17]. Also the fungal isolates were confirmed as *Aspergillus terreus* and *Aspergillus fumigatus* at Department of Pathology, Indian Agricultural Research Institute (Pusa), New Delhi, India.

The tannery effluent collected was poured into 500 mL conical flask with minimal salts medium: 3 mg/L NH₄Cl, 3 mg/L K_2HPO_4 , 5 mg/L K_2PO_4 , 1 mg/L NaCl, 1mg/L MgSO₄, covered and heated at about 80° C for 15 min and allowed to cool. The process was carried out for five (5) consecutive days in order not to denature some vital components of the effluent [18].

II.IV BIOREMOVAL OF CR FROM EFFLUENT

100 mL of tyndallized effluent with minimal salt in 250 mL Erlenmeyer flask inoculated with spore of *Aspergillus terreus* and *Aspergillus fumigatus* in separate and co-culture, at 37^oC in the orbital shaker at 100 rpm for 120 h, after every 24 h 10 mL of the effluent dawn and filtered with filter paper, then determined the Cr removed [19].

The filtrate samples of the treated effluents were centrifuged at 10,000 rpm for five (5) minutes and the supernatants were again filtered using 0.25µm micro filter membrane, in order to remove fungal biomass in it. The fungal biomass was dried in the incubator at 55°C and weight by using digital weigh balance. While residual amount of Cr in the effluent in each sample was analysed by Atomic Absorption Spectrophotometer (Model GBC 932) [20]. The percentage removal was calculated as follows:

% Removal = Initial concentration – Final concentration x100

Initial Concentration

III. RESULTS AND DISCUSSION

The tannery industries released large volume of wastewater, due to the processing of animal hide and skin to produced leather. The physicochemical parameters e.g. pH, DO, COD, BOD₅ etc of the tannery effluent shown to be reduced after treatment with individual isolates, but remarkable reduction was observed with treatment of co-culture of *Aspergillus terreus* and *Aspergillus fumigatus* table 1. This similar to the findings of Balakrishnan *et al.* [21] which reported the reduction of values of physicochemical parameters of tannery and textile effluents by microbial consortium. *Aspergillus flavus* played significant role in the reduction of TSS, TDS and BOD₅ in the tannery effluent [22]. In another finding higher DO was observed at initial stage in the textile effluent, which after treatment with the *Bacillus* and *Pseudomonas* spp for five days shows remarkable change in the BOD₅ in the effluent.

Table	1.	Dhy	reico	chan	nical	Parameters	of Tan	nary aff	luant
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Parameters	Unit	Before treatment	After treatment	
pH		9.1	7.8	
DO	mg/L	5.6	1.4	
COD	mg/L	375.1	56.3	
BOD_5	mg/L	82.5	31.2	
TSS	mg/L	262.5	46.7	
TDS	mg/L	287.3	55.2	
Turbidity	NTU	64.8	21.7	
Electrical Cond	μs/cm	10.4	4.2	

DO –Dissolved oxygen, Chemical oxygen demand, BOD- Biological oxygen demand, TSS –Total suspended solids, TDS –Total dissolved solids, Electrical conductivity.

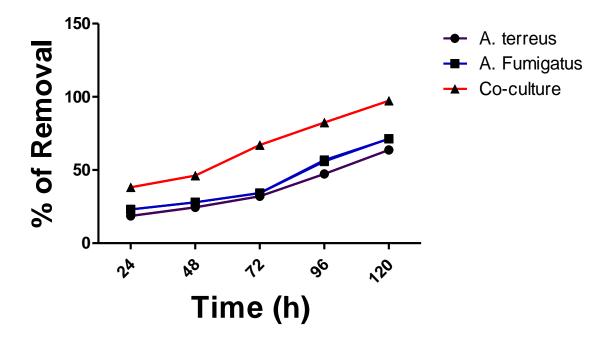


Fig1: Percentage of removal of Chromium (Cr) Aspergillus terreus, Aspergillus fumigatus and their co-culture in

120 h.

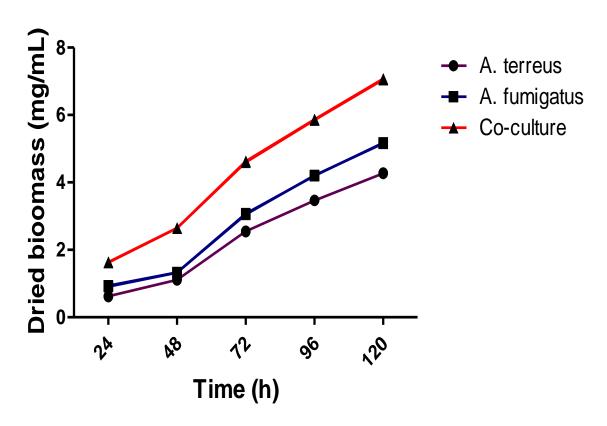


Fig. 2: Dried biomass produced by co-culture of Aspergillus terreus and Aspergillus fumigatus in

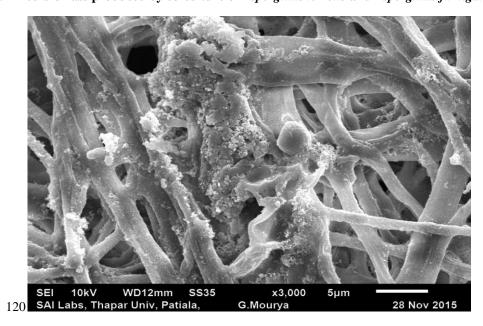


Fig. 3: Micrograph of co-culture of Aspergillus terreus and Aspergillus fumigatus treated tannery effluent

In this study, *Aspergillus fumigatus* and *Aspergillus terreus* recovered 71.34% and 63.72% within 120 h respectively. This corresponds to the findings of Murugesan *et al.*, [23] in which high amount uptake was observed by the fungal isolates at 120 h, could be attributed to time contact time needed by the fungal biomass with metal ions and their synergy effect of the fungal isolates. And the contact time have role in the metal uptake by fungal biomass, has divided into two phases: primary rapid phase which account for the major part of metal biosorption and second slow phase that contribute relatively small portion of the process. For the combination of the two fungal isolates there was optimum removal of Cr with 97.32% within 120 h. Dong *et al.*, [24] reported uptake of 96.71% of Cd from industrial effluent under agitation condition by mycelia of by mycelium of *Penicillium* spp and *Aspergillus niger*. The dried fungal biomass co-culture of *Aspergillus fumigatus* and *Aspergillus terreus* was fond to have 7.05 mg/mL; it could be attributed to the ability of the fungal isolates to with stand high concentrations of metal ions and in doing so more mycelial biomass are produced which enhanced uptake [24].

IV. CONCLUSION

Leather industries released effluents which contain heavy metals that can affect the quality of both surface and underground water. Physical and chemical methods of treatment were found insufficient, but biological treatment is found efficient and less costly for pollution control including bioremediation of heavy metals by microbes, especially fungal species which have ability to grow in wide habitat. Environmental regulation guidelines need to be strictly fallow, in order minimize pollution by industries through release of copious chemicals into the ecosystems.

REFERENCES

- [1] A. Bodalo, J.L. Gomez, E. Gomez, A.M. Hidalgo, A. Aleman, "Viability study of different reverse osmosis membranes for application in the tertiary treatment of wastes from tanning industry", Desalination, Vol. 18, Issue.4, pp.277-284, 2005.
- [2] S.G. Schrank, H.J. Jose, R.F. Moreira, H.F. Schroder, "Elucidation of the behavior of tannery wastewater under advanced oxidations", Chemosphre, Vol. 56, Issue.2, pp.411-423, 2004.
- [3] Z. Song, C.J. William, R.G. Edyvean, "Treatment of tannery wastewater by chemical coagulation", Desalination, Vol. 164, Issue.5, pp.249-259, 2004.
- [4] V.J. Sundar, R.J. Raghava, C. Muralidharan, "Cleaner Chrome tanning-emerging options", Journal of Cleaner Production, Vol. 10, Issue.3, pp.69-74, 2002.
- [5] A.R. Walsh, J.O. Halloran, "Chromium speciation in tannery effluent. II. Speciation in the effluent and in receiving estuary", Water Research, Vol. 10, Issue.4, pp.2401-2412, 1996.
- [6] S.A. Katz, H. Salem, "The biological and environmental chemistry of Chromium", VCH Publishers Inc, New York, U.S.A., 1994.
- [7] M.A. Chaudry, S. Ahmad, M.T. Malik, "Supported liquid membrane technique applicability for removal of chromium from tannery wastes", Waste Management, Vol. 17, Issue.4, pp.211-218, 1997.
- [8] B. Sarkar, "Heavy Metals in the Environment", Marcel Dekker, New York, U.S.A, 344-362, 2002.
- [9] M.I. Beydilli, S.G. Pavlostathis, W.C. Tincher, "Decolourization and toxicity screening of selected reactive azo dyes under methanogenic conditions", Water Science Technology, Vol. 38, Issue.5, pp.225-232, 1998.
- [10] M.D. Saeed, A.M. Mahmoud, "Determination of Some Physicochemical Parameters and Some Heavy Metals in Boreholes from Fagge L.G.A of Kano Metropolis Kano State Nigeria", World Journal of Analytical Chemistry, Vol. 2, Issue.2, pp.42-46, 2014.
- [11] APHA, "Standard methods for examination of water and wastewater, 20th edition, American Public Health Association, Washington, DC, U.S.A., 1998.
- [12] U. Abdullahi, A.A. Audu, Kalimullah, L. Shuaibu, "Phytoremediation of Contaminated Soils from Challawa Industrial Estate, Kano-Nigeria", Science Journal of Analytical Chemistry, Vol.4, Issue.5, pp.59-65, 2016.
- [13] A. Uzairu, O.J. Okunola, R.J. Wakawa, S.G. Adewusi, "Bioavailability Studies of Metals in Surface Water of River Challawa, Nigeria", Journal of Applied Chemistry. 2014.
- [14] M. Mohseni, F. Khosravi, M. Mohajerani, M.J. Chaichi, "Bioremediation activity of Pb (II) resistance Citrobacter sp. MKH2 isolated from heavy metal contaminated sites in Iran", Iran Journal of Science, Vol. 25, Issue.2, pp.105-110, 2014.
- [15] H.L. Devise, "A Guide to Identification of Fungi", 4th edition, ASM Press, Washington DC, U.S.A, pp.05-33, 2002.
- [16] J.P. Harley, L.M. Prescott, "Basic Laboratory and Culture Techniques", 2nd edition, WMC Brown Publisher, New York, U.S.A, Vol. 25, Issue.2, pp. 14-46, 1993.
- [17] B.G. Gabriel, (2005). Taxonomical Atlas for Identification of Fungi", 3rd edition, John Wiley publication, Florida, U.S.A., pp.11-43, 2005.
- [18] A.N. Glazer, "Microbial biotechnology concept and technologies for bioremediation in confined disposal facilities", Dredging operation and environmental research (DOER): USA Army Corps of Engineers, New York, U.S.A., pp.46-68, 1997.
- [19] L. Fude, B. Harris, M.M. Urrutia, T.J. Beveridge, "Reduction of Cr(VI) by a consortium of Sulfate reducing bacteria (SRBIII)", Appl. SEnviron. Microbiol Vol. 60, Issue.4, pp.1525-1531, 1994.
- [20] A.A. Abou Zeid, A.W. Hassanein, S.M. Hedayat, G.A. Fahd, Biosorption of some heavy metal ions, using bacterial species isolated from agricultural wastewater drains in Egypt", Journal of Applied Science and Research, Vol. 59, Issue.2, pp.372-383, 2009.
- [21] P.A. Balakrishnan, A. Arunagiri, P.G. Rao, "Ozone generation by silent electric discharge and its application in tertiary treatment of tannery effluent", Journal of Electrostatic. Vol. 56, Issue.2, pp.77-86, 2014.
- [22] H.J. Lowerence, "Basic knowledge and perspective of bioelimination of xenobiotics Compounds", J. Biotechnol, Vol. 51, Issue.4, pp.285-295, 2002.

- [23] G.S. Murugesan, K.M. Sathish, K. Swaminathan, "Arsenic from groundwater by pretreated waste tea fungal biomass", Biores. Technol, Vol. 97, Issue.8, pp.483-487, 2006.
- [24] J.L. Dong, Y.W. Zhang, R.H. Zhang, "Influence of culture conditions on laccase production and isozyme pattern in the white-rot fungus", Journal of Basic Microbiology, Vol. 45, Issue.2, pp.190-198, 2005.

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